# STRUCTURE-BIOLOGICAL ACTIVITIES RELATIONSHIPS AMONG LEUCOMYCINS AND THEIR DERIVATIVES

# SATOSHI ŌMURA, MICHIKO KATAGIRI, IWAO UMEZAWA, KANKI KOMIYAMA, TOSE MAEKAWA, KENJI SEKIKAWA,\* AKIHIRO MATSUMAE and TOJU HATA

The Kitasato Institute and Kitasato University\*, Minato-ku, Tokyo, Japan

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The relationship between chemical structure and biological activity of leucomycins and their derivatives was examined *in vivo* and *in vitro*. The biological activity of Fr-group was higher than that of Ac-group *in vitro*, but the latter showed higher blood level and lower toxicity than Fr-group. Lengthening of the carbon chain in the O-acyl group at C-4 in mycarose resulted in a marked increase in the biological activity *in vitro*, with the isovaleryl group having the highest activity. The aldehyde group in the lactone was related to antibiotic activity, but  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ -unsaturated alcohol system at C<sub>9</sub>-to C<sub>12</sub>-positions was not important.

Leucomycin, an antibiotic of the macrolide group produced in the fermentation broth of *Streptomyces kitasatoensis* was reported by HATA *et al.* in 1953<sup>1)</sup>. By further studies, we have succeeded in isolating more than 10 active substances from the leucomycin complex and the chemical structures of eight substances have been elucidated<sup>2~8)</sup>. As shown in Fig. 1, leucomycin consists of three moieties which are a 16-membered lactone, mycaminose (3, 6-dideoxy-3-dimethylamino-D-glucopyranose), and mycarose (2, 6-dideoxy-3-C-methyl-L-ribohexose).

The structural differences in the leucomycins involve the presence of hydroxyl group or O-acetyl group at the 3-position of the lactone, and the type of O-acyl group on mycarose. Leucomycin  $A_1$ ,  $A_5$ ,  $A_7$  and  $A_9$  have the hydroxyl group in the C-3 of lactone (the so-called Fr group), and  $A_3$ ,  $A_4$ ,  $A_6$  and  $A_8$  have the O-acetyl group in the same position (the so-called Ac group). The acyl groups on mycarose are isovaleryl ( $A_1$ ,  $A_3$ ), butyryl ( $A_4$ ,  $A_5$ ), propionyl ( $A_6$ ,  $A_7$ ), and acetyl ( $A_8$ ,  $A_9$ ).

This paper deals with relationship between the biological activity and the chemical structures of eight leucomycins and other related compounds which were obtained during structural studies on leucomycin.

#### Experimental

Preparations of leucomycin  $A_1$  and  $A_3 \sim A_9$  used in this study were isolated from the bulk leucomycin prepared by Toyo Jozo Ind., Co. using alumina and silica-gel column chromatography<sup>3</sup>). Diacetyl-leucomycin  $A_3$  (IX), tetrahydro-leucomycin  $A_3$  (X), hexahydroleucomycin  $A_3$  (XI), thiosemicarbazone (XII), and dehydro-leucomycin  $A_3$  (XIII) were prepared by the method reported in previous papers<sup>9~11</sup>). Isoleucomycin was obtained by treatment of leucomycin with dilute acid<sup>8</sup>). The chemical structures of these derivatives are shown in Fig. 3. VOL. XXI NO. 9

For the test of antibacterial spectrum and potency the samples were diluted with water after dissolving in a small amount of ethanol. For the determination of toxicity and blood level, the samples were diluted in 1 % sodium tartrate solution, then pH of the solution was adjusted to 4.5 with sodium hydroxide solution.

Examination of antibacterial spectrum: The minimum inhibitory concentration (MIC) of test samples were examined by an agar dilution streak method with several organisms, and the potency assay of leucomycin was done by the cup method with *B. subtilis* PCI 219 as the test bacteria.

Blood level: The leucomycins were administered to each group of 3 male mice of ddN strain by intravenous injection of 150 mg/kg (potency) via the caudal vein or by oral administration of 300 mg/kg (potency). For estimation of blood levels, blood samples were obtained from the caudal vein.

Estimation of solubility: Four mg of sample was suspended in 10 ml distilled water, and stirred for 15 minutes at 25°C. The resulting solution was filtered, the extinction of the solution was determined with a spectrophotometer at 232 m $\mu$ , and the concentration of dissolved leucomycin was calculated from the standard curve.

#### Results

### 1. Antibacterial spectrum

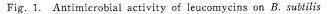
Antibacterial spectra of leucomycin  $A_1$  to  $A_9$  are given in Table 1. Each component of leucomycins strongly inhibited the growth of gram-positive bacteria but not so strongly the gram-negative bacteria. Each sample showed the same tendency in the antibacterial spectrum. As will be described later, the acetyl derivatives and thiosemicarbazone derived from leucomycin  $A_3$  also showed the same tendency. Of these eight components, their effectiveness decreased in the order of  $A_1$ ,  $A_5$ ,  $A_3$  and  $A_8$ .

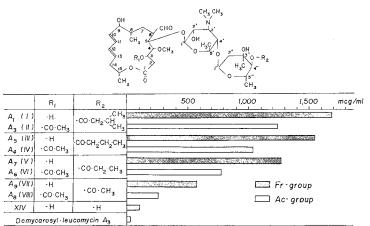
Test organism	Leucomycins (µg/ml)							
Test organism	A <sub>1</sub>	A <sub>3</sub>	A <sub>4</sub>	A <sub>5</sub>	$A_6$	$A_7$	A <sub>8</sub>	A <sub>9</sub>
Staph. aureus FDA 209P	0.04	0.15	0.15	0.80	0.30	0.15	0.60	0.30
B. subtilis PCI 219	0.30	0.60	1.25	0.30	1.25	0.30	2.50	1.25
Strept. hemolyticus	0.08	0.15	0.30	0.15	0.30	0.15	0.60	0.60
D. pneumoniae II	0.02	0.08	0.15	0.04	0.30	0.08	0.60	0.30
Coryn. diphtheriae	0.04	0.04	0.15	0.08	0.30	0.08	0.60	0.60
Neisseria gonorrhoeae	0.30	0.60	0.60	0.30	0.60	0.60	1.20	1.20
Haemophilus influenzae	0.08	0.15	0.15	0.08	0.30	0.15	0.60	0.30
Klebsiella pneumoniae PCI 602	5	10	10	5	10	10	>10	>10
S. typhimurium	10	>10	>10	10	10	>10	>10	>10
E. coli NIHJ	>10	>10	>10	>10	>10	>10	>10	>10

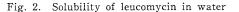
Table 1. Minimum inhibitory concentration of leucomycin components

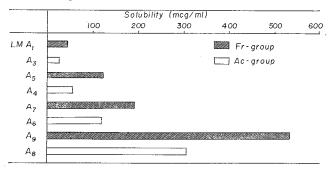
### 2. Antibacterial activity of leucomycin derivatives

The relationship between the antibacterial activity by the cup method, using *Bacillus subtilis* PCI 219 as the test organism, and the chemical structure of leucomycins is shown in Fig. 1. From these results, it is apparent that the increase in antibacterial activity of both Ac-group and Fr-group is related to an increase in the length of the carbon chain at O-acyl group on mycarose. For example,  $A_1$  and  $A_9$ have the same Fr-group, but the activity of the former is 1,690 µg/mg and that of the latter is 570 µg/mg. Deisovaleryl-leucomycin  $A_1$  (XIV) obtained from Fr-group



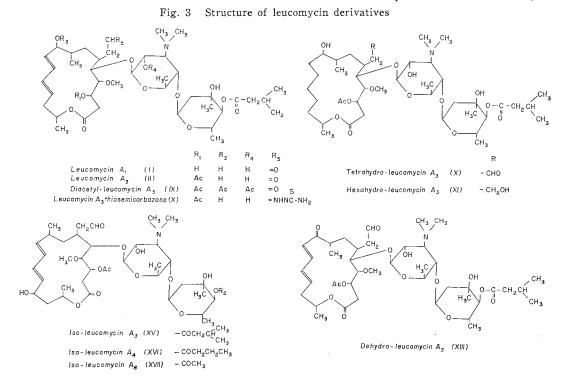






by the hydrolysis of acyl moiety, showed a remarkably low activity.

The solubilities of these samples in water are presented in Fig. 2. From the comparison of Figs. 1 and 2, it is apparent that the increase of potency of both Acgroup and Fr-group is accompanied by a decrease in solubility in water. However, acetylation of the hydroxyl group at the C-3 of lactone resulted in a decrease of both potency and solubility. It is apparent that the action of each acyl group at two positions has a different effect on the appearance of the activity.



The second se		Compounds (µg/ml)							
Test microorganism	I	II	IX	Х	XI	XII	XIII		
Staph. aureus FDA 209P	0.1	0.2	0.1	0.2	25	0.8	0.2		
Staph. albus	0.4	0.4	1.56	3.125	100	12.5	1.56		
Sarcina lutea	0.025	0.025	0.006	0.05	0.05	0.4	0.05		
B. subtilis PCI 219	0.2	0.2	0.1	0.2	50	0.8	0.2		
B. mycoides	0.4	0.4	0.4	0.4	50	6.25	0.4		
Mycobact. ATCC 607	0.05	0.1	0.1	0.1	50	0.8	1.56		
N. asteroides	1.56	1.56	1.56	1.56	100	12.5	3.125		
E. coli NIHJ	25	50	>50	>50	>100	50	100		
S. typhosa	1.56	6.25	50	>50	>100	50	100		
K. pneumoniae PCI 602	6.25	12.5	>50	12.5	>100		12.5		
V. comma (original)	0.05	0.05	0.05	0.2	12.5	0.8	0.2		

Table 2. Minimum inhibitory concentration of leucomycin derivatives

3. Structure and antibacterial activity in leucomycins and their derivatives

Minimum inhibitory concentration of leucomycins A3, A1, and their related compounds are given in Table 2, and these of iso-leuco-

Table 3. Activity of isoleuc	omycins		
Compound	Activity* (% of original)		
Iso-leucomycin A <sub>3</sub> (XV)	80		
// A4 (XVI)	90		
$H$ $A_8$ (XVII)	95		
Iso-demycarosyl leucomycin (XVIII)	83		

\* Test microorganism: B. subtilis PCI 219.

mycins<sup>8)</sup> are listed in Table 3. Increase in the degree of acetylation from  $A_1$  (I) to diacetyl- $A_3$  (IX) resulted in a slight decrease in antibacterial activity. The chemical modification of the  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ -unsaturated alcohol system,  $C_9 \sim C_{13}$  on the lactone, such as reduction to tetrahydro-leucomycin A<sub>3</sub> (X), oxidation to dehydro-leucomycin A<sub>3</sub>  $(XIII = magnamycin B^{5})$ , and acidic isomerization to isoleucomycins cause no remarkable change in the activity. From these results, it is concluded that the allylic system is not directly related to the antibacterial activity of leucomycins.

On the other hand, when the formyl group in the lactone was modified to the alcohol (XI) or thiosemicarbazone (XII), the activity was markedly decreased.

4. Toxicity

For the acute toxicity test, the compounds were administered once by intraperitoneal route to groups of 6 mice each, and the mice were observed for 14 days. The  $LD_{50}$  values in mice of each component of leucomycins are shown in Table 4. From these results, the  $LD_{50}$  values are approximately 700~800 mg/kg intraperitoneally, and

p	Ac-group					
LD <sub>50</sub> (mg/kg)		LD <sub>50</sub> (mg/kg)				
650~780	Leucomycin A <sub>3</sub> (II)	760~800 390*				
760~780	$\prime\prime$ $A_4$ (IV)	$740 \sim 750$				
700~730	// A <sub>6</sub> (VI)	770~820				
930~960	// A <sub>8</sub> (VIII)	960~970				
	Tetrahydro-leucomycin A <sub>3</sub> (X)	380*				
	Dehydro-leucomycin A <sub>3</sub> (XIII)	320*				
	650~780 760~780 700~730	$\begin{tabular}{ c c c c c c c c c c c c c c c c c c c$				

Table 4. Toxicity of leucomycins

Animal: *ddN* mice (intraperitoneal)

no marked difference is found among six components except for components  $A_8$  and  $A_9$ .

On the other hand,  $A_8$  and  $A_9$  with an acetyl group in the mycarose moiety had far less toxicity, approximately 930~970 mg/kg, than the other samples. No marked difference in toxicity was found between the Ac- and Fr-groups, but the former showed a slightly lower toxicity than the latter. The tetrahydro derivative (X) and dehydro derivative (XIII) showed a little higher toxicity than  $A_8$ .

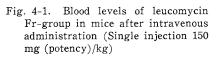
## 5. Blood level

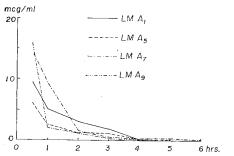
Blood level of each component of leucomycins was examined by intravenous injection or oral route.

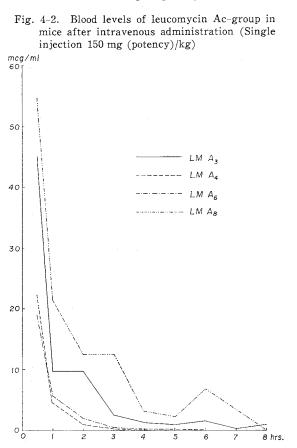
The results are shown in Fig. 4-1 and 4-2, and in Fig. 5-1 and 5-2. According to these results, the blood levels of Ac-group was several times higher than Fr-group, measurable blood levels presented for a longer time. For example, as shown in Fig. 4-1, when Fr-group was injected with a single dose of 150 mg/kg intravenously, the peak of blood level was  $10\sim15 \ \mu g/mg$  and the duration of this level was approximately  $3\sim4$  hours, while the peak of blood level of Ac-group was  $20\sim50 \ \mu g/ml$ , as shown in Fig. 4-2, and the presence of active substance in blood continued for 7 hours after the injection.

With oral administration, the highest blood level in Fr-group (Fig. 5-1) was  $2\sim4$ 

 $\mu$ g/ml and the duration of the blood level was approximately 5~7 hours after the administration. The highest blood level in Ac-group (Fig. 5-2) was  $6\sim9 \mu$ g/ml l hour after the injection, and blood levels persisted for more than 7 hours after the oral administration. Among the Ac-group, A<sub>3</sub> and A<sub>8</sub> showed a markedly higher blood level than the other samples. Thus, while the *in vitro* activity of the Ac-group is lower than that of the Fr-group,







the blood level of the former is higher than that of the latter.

### Discussion

Chemical modification of antibiotics leads to an understanding of structureactivity relationships and also may suggest methods for improving on the original compound.

The contribution of functional groups to the activity of antibiotics is complicated, making it difficult to discern structure-activity relationships. For example, in the case of streptomycin, reduction of the formyl group on streptose to hydroxyl group by catalytic hydrogenation does not change its activity<sup>12)</sup>. As shown in the present experiment, however, hydrogenation of the formyl group of leucomycin to alcohol in hexahydro derivative resulted in decrease of its activity. A result similar to the modification of the formyl group of leucomycin has been reported in studies on the chemical modification of spiramycin<sup>18)</sup>. It may be considered that the formyl group is an important group for antibacterial activity in leucomycin and the related macrolide antibiotics.

Reduction of the double bond in leucomycin, spiramycin<sup>13)</sup>, rifamycin<sup>14)</sup>, *etc.*, caused no visible change in the activity. On the other hand, reduction of the double bond in cerulenin<sup>15)</sup> and fusidic acid<sup>16)</sup> resulted considerable loss of activity. It has been reported that selective acetylation of the hydroxyl group at C-11 in the lactone C-2 in desosamine, and C-4 in oleandrose of

Fig. 5-1. Blood levles of leucomycin Fr-group in mice after oral administration (Single administration 300 mg (potency)/kg)

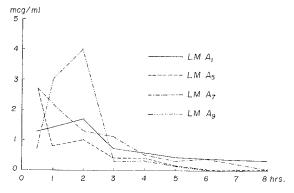
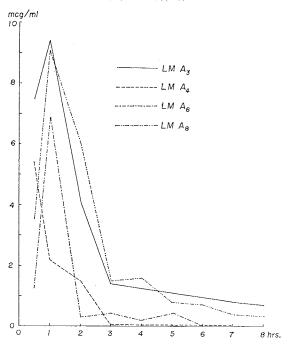


Fig. 5-2. Blood levels of leucomycin Ac-group in mice after oral administration (Single administration 300 mg (potency)/kg)



oleandomycin caused different activity changes in  $vitro^{17}$ . In addition, in the case of leucomycin, acetylation of the hydroxyl group at C-3 in the lactone decreased the activity in vitro, but that of the hydroxyl group in the mycarose resulted in increased activity.

Increased activity was obtained with leucomycins by lengthening the aliphatic chain of the acyl group attached to C-4 of the mycarose, but the reverse is true with the Oacyl group at C-16 of fusidic acid<sup>16</sup>.

It has been known that the activity of macrolide antibiotics such as erythromycin<sup>18)</sup>, spiramycin<sup>19)</sup>, and oleandomycin<sup>17)</sup> was increased *in vivo* by acetylation. As shown in the present experiment *in vivo*, the results with the Fr- and Ac-groups of leucomycin were the same as that of other macrolides which show an increasing activity with acylation. Though it has been reported that leucomycin shows a relatively low blood level and shorter duration in blood<sup>20~21)</sup>, it is considered that these results were due to the use of the Fr-group, especially  $A_1$ .

#### References

- HATA, T.; Y. SANO, N. OHKI, Y. YOKOYAMA, A. MATSUMAE & S. ITO: Leucomycin, a new antibiotic. J. Antibiotics, Ser. A 6: 87~89, 1953
- WATANABE, T.; H. NISHIDA, J. ABE & K. SATAKE : Studies on leucomycin. III. Isolation and properties of six antibacterial components in leucomycin complex. Bull. Chem. Soc. Japan 33 : 1104~1108, 1960
- HATA, T.; S. OMURA, M. KATAGIRI, K. MIYAZAKI, S. SATO & K. TSUCHIDA : The chemistry of leucomycin. V. Isolation of new components from leucomycin complex. J. Antibiotics (to be published)
- OMURA, S.; H. OGURA & T. HATA: The chemistry of the leucomycin. I. Partial structure of leucomycin A<sub>3</sub>. Tetrahedron Letters No. 7: 609~613, 1967
- 5) OMURA, S.; H. OGURA & T. HATA: The chemistry of the leucomycins. II. Structure and stereochemistry of leucomycin A<sub>3</sub>. Tetrahedron Letters No. 14: 1267~1271, 1967
- 6) OMURA, S.; M. KATAGIRI & T. HATA: The chemistry of leucomycins. IV. Structure of leucomycin A<sub>1</sub>. J. Antibiotics 21: 199~203, 1968
- 7) OMURA, S.; M. KATAGIRI & T. HATA: The chemistry of leucomycins. VI. Structures of leucomycin A<sub>4</sub>, A<sub>5</sub>, A<sub>6</sub>, A<sub>7</sub>, A<sub>8</sub> and A<sub>9</sub>. J. Antibiotics 21: 272~278, 1968
- 8) OMURA, S.; M. KATAGIRI, K. NAYA and T. HATA: Absolute configuration of leucomycin A<sub>3</sub>. Presented at the 25th Annual Meeting of the Pharmaceutical Society of Japan, Tokyo, April 1968
- 9) OMURA, S.; M. KATAGIRI, H. OGURA & T. HATA: The chemistry of leucomycins. I. Partial structure of leucomycin A<sub>3</sub>. Chem. Pharm. Bull. (Tokyo) 15: 1529~1533, 1967
- 10) OMURA, S.; M. KATAGIRI, H. OGURA & T. HATA: The chemistry of leucomycins. II. Glycosidic linkages of mycaminose and mycarose on leucomycin A<sub>3</sub>. Chem. Pharm. Bull. (Tokyo) 16: 1167~1173, 1968
- OMURA, S.; M. KATAGIRI, H. OGURA & T. HATA: The chemistry of leucomycin. III. Structure and stereochemistry of leucomycin A<sub>3</sub>. Chem. Pharm. Bull. (Tokyo) 16:1181~1186, 1968
- KAVANAGH, F.: Activities of twenty-two antibacterial substances against nine species of bacteria. J. Bacteriol. 54: 761~766, 1947
- ADAMSKI, R. J.; H. HEYMANN, S. G. GEFTIC & S. S. BARKULIS: Preparation and antibacterial activity of some spiramycin derivatives. J. Med. Chem. 2:932~934, 1966
- 14) SENSI, P.; N. MAGGI, S. FÜRESZ & G. MAFFII: Chemical modifications and biological properties of rifamycins. Antimicr. Agents & Chemoth. 1966: 699~714, 1967
- 15) Ната, Т., S. Óмика : Unpublished work. cf. S. Óмика, М. Катадікі, А. Nakagawa, Y. Sano, S. Nomuka & T. Hata : Studies on cerulenin. V. Structure of cerulenin. J. Antibiotics, Ser. A 20 : 349~354, 1967
- 16) GODTFREDSEN, W. O.; C. ALBRETHSEN, W. V. DAEHNE, L. TYBRING & S. VANGEDAL: Transformations of fusidic acid and the relationship between structure and antibacterial activity. Antimicr. Agents & Chemoth. 1965: 132~137, 1966
- CELMER, W. D. : Triacetyloleandomycin: Biological correlations. Antibiot. Ann. 1958/1959 : 277 ~283, 1959
- STEPHENS, V. C. & J. W. CONINE : Esters of erythromycin. III. Esters of low molecular weight aliphatic acids. Antibiot. Ann. 1958/1959 : 346~353, 1959
- 19) TAKAHIRA, H.; H. KATŌ, N. SUGIYAMA, S. ISHII, T. HANEDA, K. UZU, K. KUMABE & R. KOJIMA : Fundamental studies on acetyl spiramycin. J. Antibiotics, Ser. B 19:95~100, 1966
- 20) KANAMORI, K. : The blood level and the distribution in tissues of leucomycin. II. Tissue concentrations after oral administration of leucomycin. Chemotherapy 5: 87~90, 1957
- HOSHINO, Y.; I. UMEZAWA, H. YAMAMOTO, A. MATSUMAE & T. HATA: Studies on leucomycin A<sub>1</sub>.
  II. Studies on blood and tissue levels, and toxicities. J. Antibiotics, Ser. A 19: 30~36, 1966